

For In Vitro Diagnostic Use

Mouse/Rabbit PolyDetector AEC HRP Red Detection System

Intended Use

For In Vitro Diagnostic Use.

Summary and Explanation

The **Mouse/Rabbit PolyDetector AEC HRP Red System** is a non-biotin, 2-step polymeric detection system that allows for the demonstration of antigens in paraffin-embedded tissue, cryostat sections, blood smears, cytosmears, and cell preparations. The PolyDetector kits have been developed using a proprietary tandem hyperlabelling technology used to directly label immunoglobulins with enzymes. This ensures consistent and reproducible immunostaining for all types of nuclear, cytoplasmic and membranal antigens, in different types of tissues.

The increased sensitivity of the **Mouse/Rabbit PolyDetector AEC HRP Red Detection System** allows for rapid staining procedures without compromising stain quality. The **Mouse/Rabbit PolyDetector AEC HRP Red Detection System** is suitable for use with mouse IgG and IgM and rabbit primary antibodies, both monoclonal and polyclonal. The **Mouse/Rabbit PolyDetector AEC HRP Red Detection System** kits are optimized for use with Bio SB primary antibodies; however, they are universal kits and therefore work equally well with prediluted and concentrated antibodies from different vendors.

Presentation

The **Mouse/Rabbit PolyDetector AEC HRP Red Detection System** contains a Peroxidase Blocker solution, an Anti-Mouse/Rabbit Horseradish Peroxidase solution, and an AEC Chromogen solution. All the components are buffered with stabilizers and an anti-microbial.

Availability	
Catalog Number	Volume
BSB 0202S	5 mL Each
BSB 0202	15 mL Each
BSB 0204	50 mL Each
BSB 0206	100 mL Each
BSB 0208	200 mL Each
BSB 0208A	1000 mL Each

Storage: Store at 2°C - 8°C

Stability: Stable up to the expiration date listed on the label. Do not use this product after the expiration date listed on the product label.

Protocol

Preparation of Working Solutions

The **PolyDetector Peroxidase Blocker**, the **Anti-Mouse/Rabbit Horseradish Peroxidase Label**, and the **AEC Chromogen Solution** are ready-to-use working solutions and require no further preparation.

Mounting Protocol

A. AquaMounter Protocol

1. After the histological, immunohistochemical or in situ hybridization staining procedure is completed, rinse slides in deionized water. Do not incubate tissue or cell specimens in solvents such as alcohol, toluene, or xylene.
2. After counterstaining, apply 1-3 drops of AquaMounter (BSB 0090- BSB0093) to each slide making sure the specimen is covered.
3. Coverslip.
4. Observe under a light microscope.

B. ChromoProtector Permanent Mounting Protocol

1. After the histological, immunohistochemical or in situ hybridization staining procedure is completed, rinse slides in deionized water. Do not incubate tissue or cell specimens in solvents such as alcohol, toluene, or xylene.
2. Using a Coplin jar or a staining dish, immerse slides with tissues in ChromoProtector (BSB 0151 – BSB 0156) or lay wet slides horizontally and apply sufficient drops of ChromoProtector (BSB 0151 – BSB 0156) to completely cover the tissue. Carefully spread ChromoProtector if needed, but avoid contacting the tissue.
3. Incubate slides for ten minutes at 60 °C to allow ChromoProtector to penetrate tissues.
4. Remove excess ChromoProtector by placing slides vertically over an absorbent material and let excess drain off into absorbent material. Do not rinse slides.

5. Allow slides to COMPLETELY air dry.
6. NOTE: The ChromoProtector will protect tissue from drying artifacts during the air-drying process.
7. When slides are completely dried, they can be mounted using most standard mounting methods such as aqueous or permanent.
8. Organic solvent permanent mounting medium such as PermaMunter (Cat# BSB 0094-0097) or similar permanent mounting media, or the organic solvent-free XyGreen PermaMunter (Cat # BSB 0169-0174), can be added directly to the slide until the tissue or cell specimen is covered.
9. If the permanent mounting medium does not spread evenly on the dry slide, the slide can be dipped in toluene or xylene for 1 – 2 seconds to aid spreading of the mounting medium. Use a minimum amount of mounting medium so that slides dry rapidly.
10. Apply coverslip and air dry before microscopic observation.

Recommended Immunohistochemical Protocol

1. Cut and mount 3-4 micron formalin-fixed paraffin-embedded tissues on positive charged slides.
2. Air dry for 2 hours at 58° C.
3. Deparaffinize, dehydrate and rehydrate tissues.
4. Subject tissues to heat epitope retrieval using a suitable retrieval solution such as ImmunoDNA Retriever with Citrate (BSB 0020-BSB 0023) or EDTA (BSB 0030-BSB 0033).
5. Wash with 5 changes of IHC Wash buffer.
6. Place slides in **PolyDetector Peroxidase Blocker** for 5 min.
7. Wash with 3 changes of IHC wash buffer.
8. Cover tissue with the **Primary Antibody** following manufacturer's recommended protocol. If using concentrated antibodies, we suggest using our **ImmunoDetector Protein Blocker/Antibody Diluent** to dilute antibodies.
9. Wash with 3 changes of IHC wash buffer.
10. Cover tissue with **PolyDetector HRP Label**, incubate for 45 min.
11. Rinse with 3 changes of IHC wash buffer.
12. Cover tissue with **AEC HRP Red Chromogen**, incubate for 10 minutes.
13. Rinse with 5 changes of DI water
14. Counterstain.
15. Coverslip with a water-based mounting medium.









Abbreviated Immunohistochemical Protocol	
Step	PolyDetector HRP
Peroxidase Blocker	5 minutes
Primary Antibody	45 - 60 minutes
HRP Label	45 minutes
DAB Substrate-Chromogen	10 minutes
Counterstaining	Time varies with counterstain

Precautions

1. For professional users only. Results should be interpreted by a medical professional.
2. Ensure proper handling procedures are used with reagent. Always wear proper personal protective equipment such as laboratory coat, goggles and gloves when handling reagents
3. This product contains components with sodium azide (NaN₃), a toxic chemical. At product concentrations it is not classified as hazardous due to its low concentration. Sodium azide may react with plumbing to form highly explosive build-ups of metal azides. Upon disposal, flush with large volumes of water to prevent metal azide build up.
4. Minimize microbial contamination of reagents
5. Dispose of unused solution according to local and federal regulations.
6. Do not ingest reagent. If reagent ingested, contact a poison control center immediately.
7. Avoid contact with eyes. Flush with large quantities of water if contact occurs
8. Follow safety precautions for the heating device (TintoRetriever Pressure Cooker or similar).
9. For complete recommendations for handling biological specimens please refer to the CDC document, "Guidelines for Safe Work Practices in Human and Animal Medical Diagnostic Laboratories" (1).

References

1. U.S. Department of Health and Human Services: Centers for Disease Control and Prevention. Guidelines for Safe Work Practices in Human and Animal Medical Diagnostic Laboratories. Supplement / Vol. 61, January 6, 2012.

Symbol Key / Légende des symboles/Erläuterung der Symbole			
		Storage Temperature Limites de température Zulässiger Temperaturbereich	
		Read Instructions for Use Consulter les instructions d'utilisation Gebrauchsanweisung beachten	
			
In Vitro Diagnostic Medical Device Dispositif médical de diagnostic in vitro In-Vitro-Diagnostikum			Manufacturer Fabricant Hersteller
			Expiration Date Utiliser jusque Verwendbar bis
			Catalog Number Référence du catalogue Bestellnummer
			Lot Number Code du lot Chargenbezeichnung



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